Dihydroramulosin from Botrytis sp.

Donald B. Stierle,* Andrea A. Stierle, and Anita Kunz

Department of Chemistry, Montana Tech of the University of Montana, Butte, Montana 59701

Received July 18, 1997

Botrytis sp., isolated from the inner bark of the Pacific yew, *Taxus brevifolia*, was shown to produce ramulosin (1), 6-hydroxyramulosin (2), and the new compound 8-dihydroramulosin (3). The structure of dihydroramulosin was deduced from the NMR spectra and confirmed by chemical conversion from ramulosin.

For the past five years we have been looking at the endosymbionts found in the inner bark of the Pacific yew, *Taxus brevifolia.*¹ In the course of these investigations we have isolated a variety of fungi. The chloroform extract of one of these fungal growths showed strong antibiotic activity against *Staphylococcus aureus, Candida albicans,* and *Vibrio harveyii,* so we decided to look at the bioactive components. The fungus, *Botrytis* sp. was grown in mycological broth for 21 days and extracted with CH₂Cl₂. This extract had strong antibiotic activity. The extract was fractionated on LH-20 and then by counter current column chromatography (CCCC). The antibiotic activity against *S. aureus* was followed during these fractionations to give three antimicrobial compounds. These compounds showed moderate antimicrobial activity in the disk assay.

The two less polar antimicrobial compounds were identified as ramulosin (1) and 6-hydroxyramulosin (2) by comparison of their NMR spectra to those of the known compounds.^{2,3} These two fungal metabolites have previously been isolated from *Pestalosia ramulosa*,^{2,3} and ramulosin has been more recently reported from *Truncatella hartigii*⁴ and *Hypoxylon howeianum*.⁵ The absolute configuration of ramulosin has also been established as shown in 1.⁶ Because the ¹³C NMR data have not been reported for these compounds, they are shown in Table 2.

The more polar antimicrobial metabolite, **3**, gave a very weak parent ion in the EIMS at m/z 184, and larger fragment ions at m/z 169 (M⁺–CH₃) and 166 (M⁺–H₂O). The molecular formula of C₁₀H₁₆O₃ was deduced from the HRCIMS, which gave a strong [M⁺ + 1] peak at m/z 185. Alcohol **3** readily formed a monoacetate when treated with Ac₂O/pyridine. These data suggested that **3** was a dihydroramulosin. Detailed examination of the 500 MHz ¹H NMR and COSY spectra of **3** suggested the alcohol functionality was at C-8. Reduction of the enol group in ramulosin should then give **3**.

Reduction of the enol of ramulosin proved to be not as simple as originally thought. When ramulosin was treated with sodium borohydride for 10 min at 20 °C and acetylated with Ac₂O-pyridine the product was found to be a mixture of diacetates. These diacetates were formed from the reduction of both the enol and ester groups. The diacetates were separated by HPLC on Si gel. One of the acetoxy protons for the more polar major isomer, **6**, appeared as a doublet (J = 8.5 Hz) at δ 5.54. This proton was assigned to C-1 and given the relative stereochemistry shown based on the large diaxial proton coupling. The second acetoxy proton appeared as a doublet of triplets (J = 10.2, 4.6 Hz)

Table 1. ¹H NMR Data for Compounds 1-4 (500 MHz, CDCl₃)

C no.	1	2	3	4
3	4.43 (m)	4.50 (m)	4.39 (m)	4.41 (m)
4	1.90 (m)	1.89 (dm, 13.4)	2.23 (ddd,	2.08 (m)
			14.1, 6.4, 3.8)	
	1.28 (m)	1.29 (m)	1.23 (m)	1.32 (m)
4a	2.48 (tm,	2.89 (br t, 11.1)	2.13 (m)	2.26 (m)
	11.2)			
5	2.34 (m)	1.97 (dm, 13.1)	1.61 (br d, 12.7)	1.63 (m)
	1.12 (m)	1.35 (m)	0.92	1.20 (m)
			(qd, 13.0, 3.2)	
6	1.88 (m)	4.34 (br s)	1.7 (m)	2.08 (m)
	1.85 (m)		1.23 (m)	1.36 (m)
7	1.62 (m)	2.63 (br d, 19.3)	1.87 (dt, 13.0, 4.0)	2.01 (m)
	1.90 (m)	2.39 (d, 19.3)	1.72 (m)	1.70 (m)
8			3.61 (dt, 11.3, 5.2)	4.92 (br s)
8a			2.95 (br t, 5.3)	3.03 (br t,
				4.8)
CH_3	1.35 (d,	1.38 (d, 6.2)	1.31 (d, 6.2)	1.32
	6.2)			(d, 6.2)
Ac				2.05 (s,
				3H)

Table 2.¹³C NMR Data for Compounds 1–3

C no.	1	2	3
1	174.7 (s)	171.9	174.9 (s)
3	76.5 (d)	76.8	73.4 (d)
4	37.5 (t)	37.5	36.5 (t)
4a	32.9 (d)	26.4	31.9 (d)
5	29.5 (t)	36.9	30.9 (t)
6	29.0 (t)	63.8	23.4 (t)
7	20.9 (t)	35.8	32.8 (t)
8	171.8 (s)	171.5	70.1 (d)
8a	96.8 (s)	96.5	44.2 (d)
CH_3	21.7 (q)	21.7	20.6 (q)

at δ 4.72 and was assigned at C-8 and given the relative stereochemistry shown, again based on the coupling. Reduction of the ester group with sodium borohydride gave a surprising result. When ramulosin was treated with sodium borohydride at 0 °C, the reaction gave only one product that had an identical ¹H NMR spectrum to that of dihydroramulosin (**3**).

The relative stereochemistry of **3** was established from difference NOE studies. Irradiation of the proton at δ 4.39 (H-3) gave a strong enhancement at δ 2.95 (H-8a), which indicated that these protons had a cis arrangement. Irradiation at δ 2.95 gave the expected enhancement at δ 4.39 and also enhanced the other bridgehead proton δ 2.13 (H-4a). The other irradiations and enhancements are shown in Figure 1.

^{*} To whom correspondence should be addressed. Tel.: (406) 496-4717. Fax: (406) 496-4688. E-mail: DSTIERLE@MTECH.EDU.

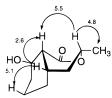
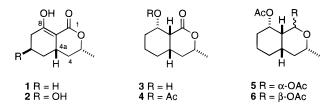


Figure 1. Difference NOE experiments on **3** (numbers on the arrows indicate the % NOE enhancements).



Experimental Section

General Experimental Procedures. ¹H and ¹³C NMR spectra were recorded in CDCl₃ on Bruker DRX-500 and DRX-250 spectrometers. ¹H NMR spectra were recorded at 500 MHz, and the ¹³C NMR spectra were recorded at 67.5 MHz. All of the chemical shifts were recorded with respect to the deuterated solvent shift. The IR spectrum was recorded on a Perkin–Elmer 1310 spectrometer. The optical rotation was recorded on a Perkin–Elmer 241 MC polarimeter using a 1-mL cell. The melting points were recorded on a Mel-TempII melting apparatus. The MS were provided by the Montana State Mass Spectrometer facility at Montana State University. All solvents used were spectral grade.

Extraction and Isolation. The isolation of this fungus has been described previously.¹ The identification of this isolate as *Botrytis* sp. was confirmed by fatty acid analysis through Microbial ID Inc. (ID 2377). *Botrytis* sp. was grown in mycological broth (2×2 L) in still culture. After 21 days, the mycelia was filtered off and the broth extracted with CH₂-Cl₂ (3 X with 1 L of CH₂Cl₂) to give 0.3076 g of a brown oil. This crude extract was fractionated on LH-20 (CHCl₃-MeOH 1:1). The fractions that showed antibiotic activity against *S. aureus* were further fractionated by CCCC using hexane–EtOAc–MeOH–H₂O (50:70:80:65) to give three major bioactive fractions.

Ramulosin (1): white crystals; mp 115–117 °C, $[\alpha]^{23}_{\rm D}$ +14.5° (*c* 0.5 MeOH), which exhibited comparable spectral (¹H NMR) data to published values.²

6-Hydroxyramulosin (2): white solid; mp 122–125 °C, $[\alpha]^{23}_{D}$ +48.9° (*c* 0.47 MeOH), which exhibited comparable spectral (¹H NMR) data to published values.²

Dihydroramulosin (3): white solid; mp 142–144 °C, $[\alpha]^{23}_{D}$ +18.6° (*c* 0.59 MeOH); ¹H NMR data, see Table 1; ¹³C NMR data, see Table 2; EIMS (70 eV) *m*/*z* 184 (1), 169 (2), 166 (3),

Table 3. ¹H NMR Data for 5, and 6, Selected Peaks

C no.	5	6
1	6.15 (d, 3.0)	5.54 (d, 8.5)
3	3.91 (m)	3.68 (pd, 6.2, 4.2)
8	4.61 (td, 10.7, 4.6)	4.72 (dt, 10.2, 4.6)
CH3	1.12 (d, 6.2)	1.18 (d, 6.2)
OAc's	2.03, 1.97	2.05, 1.98

156 (71), 113 (100), 95 (50), 80 (54); HRCIMS m/z 185.1174 (calcd for C₁₀H₁₇O₃, 185.1178).

Acetylation of 3. Compound **3** (3 mg) was dissolved in Ac_2O (1.0 mL) and pyridine (1.0 mL) and the solution stirred. After 24 h the volatiles were removed in vacuo and the acetate purified by flash Si gel chromatography, giving acetoxydihydroramulosin **(4)**: oil; ¹H NMR data, see Table 1; EIMS (70 eV) m/z M⁺ not observed, 166 [M⁺ – AcOH] (4), 80 (100), 79 (44).

Reduction and Acetylation of Ramulosin. Ramulosin (5 mg) was dissolved in 3 mL of MeOH, NaBH₄ (5 mg) was added, and the reaction was stirred for 10 min at 20 °C. The MeOH was removed from the reaction solution in vacuo and H₂O (3 mL) and CH₂Cl₂ (3 mL) added. The organic layer was separated, dried (Na₂SO₄), and the crude products acetylated by placing them in Ac₂O (1.0 mL) and pyridine (1.0 mL). After 24 h, the volatiles were removed in vacuo and the diacetates separated by HPLC on Si gel (Rainin), to give the less polar diacetate (5): (¹H NMR data, see Table 3) and the more polar diacetate (6): (¹H NMR data, see Table 3).

Reduction of Ramulosin at 0 °C. Ramulosin (5 mg) was dissolved in 3 mL of MeOH, NaBH₄ (5 mg) was added, and the reaction was stirred for 5 min at 0 °C. The MeOH was removed from the reaction solution in vacuo, and H_2O (3 mL) and CH_2Cl_2 (3 mL) were added. The organic layer was separated and dried (Na₂SO₄). The major product was purified by HPLC on Si gel (Rainin) to give a compound that exhibited a ¹H NMR spectrum identical to that of dihydroramulosin (**3**).

Acknowledgment. The authors would like to thank Scott Busse of the Department of Chemistry, Montana State University, for NMR technical help. The NMR facility was supported by NSF grant 9506620.

References and Notes

- Stierle, A.; Strobel, G.; Stierle, D. B.; Grothaus, P.; Bignami, G. J. Nat. Prod. 1995, 58, 1315–1324.
- (2) Stodola, F. H.; Cabot, C.; Benjamin, C. R. *Biochem. J.* **1964**, *93*, 92– 97.
- (3) Tanenbaum, S. W.; Agarwal, S. G.; Williams, T.; Pitcher, N. Tetrahedron Lett. 1970, 27, 2377–2380.
 (4) Turner, W. B.; Aldridge, D. C. Fungal Metabolites II; Academic: New View Construction of the second s
- York, 1983; p 87. (5) Turner, W. B.; Aldridge, D. C. *Fungal Metabolites II*; Academic: New
- York, 1983; p 507. (6) Findlay, J. A.; Matsoukas, J. M.; Krepinsky, J. *Can. J. Chem.* **1976**,

54, 3419–3423.

NP9703407